



First Detection of FOX-1 AmpC β -lactamase Gene Expression Among *Escherichia coli* Isolated from Abattoir Samples in Abakaliki, Nigeria

Ejikeugwu Chika^{1*}, Esimone Charles², Iroha lfeanyichukwu¹ and Adikwu Michael³

¹Department of Applied Microbiology, Faculty of Science, Ebonyi State University, Abakaliki, Nigeria

²Department of Pharmaceutical Microbiology and Biotechnology, Faculty of Pharmaceutical Sciences, Nnamdi Azikiwe University, Awka, Nigeria

³Department of Pharmaceutics, University of Nigeria Nsukka, Nsukka, Nigeria

ARTICLE INFO

Article history:

Received: 21 September 2017

Accepted: 8 February 2018

Online:

DOI 10.5001/omj.2018.44

Keywords:

Abattoirs; Cephalosporinase; Antibiotic Resistance; *Escherichia coli*; FOX-1 genes; Multidrug Resistance Bacteria; Nigeria.

ABSTRACT

Objectives: Gram-negative bacteria represent the most relevant reservoir of resistance to antibiotics in the environment. The natural selection of resistant clones of bacteria in the environment by antimicrobial selective pressure is a relevant mechanism for spreading antibiotic resistance traits in both the community and hospital environment. This is in scenarios where antimicrobials are used irrationally, and even in the propagation of livestock, poultry birds, and for other veterinary purposes. This study sought to detect the prevalence of FOX-1 AmpC β -lactamase genes from abattoir samples. **Methods:** The isolation of *Escherichia coli*, antimicrobial susceptibility testing, and β -lactamase characterization was carried out using standard microbiology techniques. The production of AmpC β -lactamase was phenotypically carried out using the ceftaxime-cloxacillin double-disk synergy test (CC-DDST), and FOX-1 AmpC genes was detected in the *E. coli* isolates using multiplex polymerase chain reaction. **Results:** Forty-eight *E. coli* isolates were recovered from the anal swabs of cows and 35 (72.9%) isolates were positive for the production of β -lactamase. Notably, high percentages of resistance to ceftaxime (91.7%), ceftriaxone (83.3%), imipenem (85.4%), ceftazidime (87.5%), ofloxacin (81.3%), and gentamicin (85.4%) were found. FOX-1 genes were detected in three (6.3%) of the 48 *E. coli* isolates phenotypically screened for AmpC enzyme production. **Conclusions:** Abattoirs could represent a major reservoir of resistance genes especially AmpC β -lactamase, and this could serve as a route for the dissemination of multidrug-resistant bacteria in the community. Thus, the molecular identification of drug-resistant genes is vital for a reliable epidemiological investigation and the forestalling of the emergence and spread of these organisms through the food chain in this region.

In most parts of the world, especially developing countries including Nigeria, the detection of resistant genes and/or multidrug-resistant (MDR) enzymes responsible for the negative response of pathogenic bacteria to potent antimicrobial onslaught is still ill-detected in our hospitals. This is because routine antimicrobial susceptibility studies (as is obtainable in most of our hospitals) is ineffective in detecting these MDR organisms. Gram-negative bacteria represent the most relevant reservoir of resistance to antibiotics in the environment. The natural selection of resistant clones of bacteria in the environment

by antimicrobial selective pressure is a relevant mechanism for spreading antibiotic resistance traits in the community and hospital environment.^{1,2} This is in scenarios where antimicrobials are used irrationally, and even in the propagation of livestock, poultry birds, and other veterinary purposes. AmpC β -lactamases are cephalosporinases that belong to molecular class C β -lactamases and are encoded on the chromosomes of many gram-negative bacteria.¹⁻³ The genes encoding AmpC β -lactamases are much more frequently chromosomal than plasmid-mediated. The AmpC β -lactamases are clinically important β -lactamases because they confer antimicrobial

*Corresponding author: ✉ejikeugwu_chika@yahoo.com

resistance to the narrow-spectrum, expanded-spectrum, and the broad-spectrum cephalosporins including cefotaxime, ceftazidime, ceftriaxone, aztreonam, and the penicillins. Resistance is also expressed towards β -lactamase inhibitors such as amoxicillin-clavulanic acid.^{3,4} Most of the genera of the family *Enterobacteriaceae* produce AmpC enzymes through an inducible mechanism in which case the presence of broad-spectrum antibiotics sparks enzyme production in the organism.^{4,5} The production of some MDR enzymes such as extended-spectrum beta-lactamases (ESBLs) by gram-negative bacteria can be masked by the coexpression of AmpC in the same organism.^{6,7} One of the biggest challenges facing the health sector across the globe, especially in the area of infection control and prevention, is in the adequate containment of multidrug-resistant gram-negative organisms (MDRGNOs). It is worrisome that MDRGNOs are not restricted to the hospital environment and bacterial pathogens including *Klebsiella* species, *Pseudomonas aeruginosa*, and *Escherichia coli*, which produce enzymes that degrade some of the world's potent and effective drugs such as carbapenems and cephalosporins, are now found in the community.⁸⁻¹² Measuring the prevalence of these organisms in both hospital and community settings coupled with proper use of available drugs and susceptibility testing of these organisms before the prescription or use of some expanded-spectrum drugs (especially in hospital environments) is critical to reducing the possible risks associated with infections due to MDRGNOs.⁶⁻¹²

Food-producing animals harboring MDR genes together with genes that mediate the production of some high-profile antibiotic hydrolyzing enzymes such as AmpC enzymes possess health risks to the human population due to their potential contribution in the spread of drug-resistant microorganisms in the community. We used multiplex polymerase chain reaction (PCR) to detect the occurrence of FOX-1 genes in *E. coli* isolates from abattoir sources in Abakaliki, Nigeria.

METHODS

In line with the World Medical Association Declaration of Helsinki on the principles for medical research involving human subjects and identifiable human and animal material/data, this research was undertaken based on a thorough

knowledge of the scientific literature and adequate laboratory protocol. Oral consent was sought from the abattoir operators for the collection and processing of the anal swab samples. There was no written ethical approval as the samples were collected at the point of slaughter, and the animals were handled with respect and total care. There was minimal pain incurred by the animals during sample collection.

A total of 120 samples were collected from the anal region of cows in a local abattoir in Abakaliki, Nigeria, using sterile swab sticks. To isolate *E. coli*, a loopful of the turbid solution from overnight nutrient broth culture was plated onto eosin methylene blue (EMB) agar and MacConkey agar (Oxoid, UK) plates. Each plate was incubated at 30 °C for 18 hours and showed a positive reaction. Suspect colonies of *E. coli* was aseptically subcultured onto freshly prepared EMB and MacConkey agar plates for the isolation of pure cultures, and these were incubated at 30 °C for 18 hours. *E. coli* was identified using standard microbiology identification technique.⁸

Antimicrobial susceptibility studies were carried out using the modified Kirby-Bauer disk diffusion method on Mueller-Hinton (MH) agar plates (Oxoid, UK) as previously described.^{9,10} Briefly, the test antibiotic disks were aseptically placed at a distance of 15 mm apart on MH agar plates already swabbed or inoculated with the *E. coli* isolates (adjusted to 0.5 McFarland turbidity standards). Each plate was incubated at 30 °C for 18 hours. *E. coli* ATCC 25922 strain was used as the quality control strain for the antimicrobial susceptibility test. Inhibition zones were interpreted according to Clinical and Laboratory Standard Institute (CLSI) criteria.

To characterize β -lactamase the impregnated end of the nitrocefin stick was used to pick the test isolate; and the stick was macroscopically observed after 5–15 minutes for a color change.¹¹ A positive reaction was shown by the development of a pink-red color while a negative reaction was indicated by no color change.

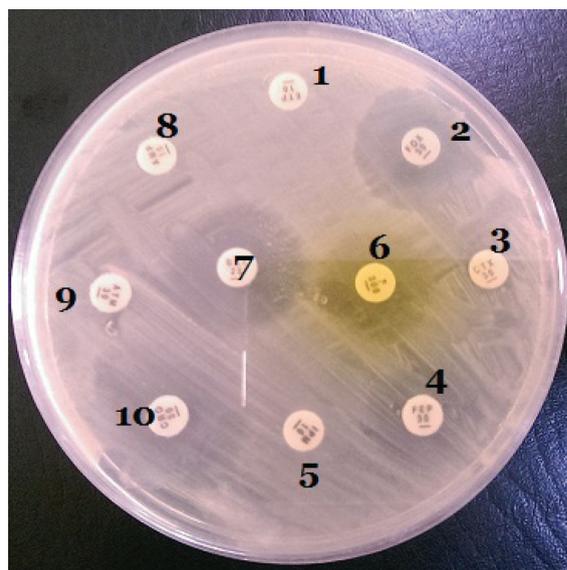
Bacterial strains that produce AmpC β -lactamase are resistant to the cephamycins but susceptible to the fourth generation cephalosporin, cefepime.^{12,13} All test isolates were subjected to the antimicrobial activity of cefoxitin disk (30 μ g) on aseptically streaked MH agar plates. Each plate was incubated at 30 °C for

Table 1: Frequency of *Escherichia coli* from anal swab samples.

Sample source	Number of samples	<i>E. coli</i> , n (%)	β -lactamase production, n (%)
Anal swabs of cow	120	48 (40.0)	35 (72.9)

18 hours.^{10,12} Inhibition zones were interpreted according to CLSI criteria.

Cefoxitin-cloxacillin double-disk synergy test (CC-DDST) was performed according to the method described by Polsfuss et al.¹⁴ Single disks containing 30 μ g of cefoxitin were placed 20 mm away from a disk containing 200 μ g of cloxacillin on MH agar plates already inoculated with the test bacteria (equivalent to 0.5 McFarland turbidity standards). Each plate was incubated at 30 °C for 18 hours. Inhibition zones were interpreted according to CLSI criteria. A difference of 4 mm in the cefoxitin-cloxacillin inhibition zones minus the



1: Ertapenem (10 μ g); 2: Cefoxitin (30 μ g); 3: Cefotaxime (30 μ g); 4: Ciprofloxacin (10 μ g); 5: Imipenem (10 μ g); 6: Nitrofurantoin (10 μ g); 7: Meropenem (10 μ g); 8: Ampicillin (10 μ g); 9: Aztreonam (30 μ g); 10: Ceftriaxone (30 μ g).

Figure 2: Susceptibility test plate of an *Escherichia coli* isolate cultured on Mueller-Hinton agar plate.

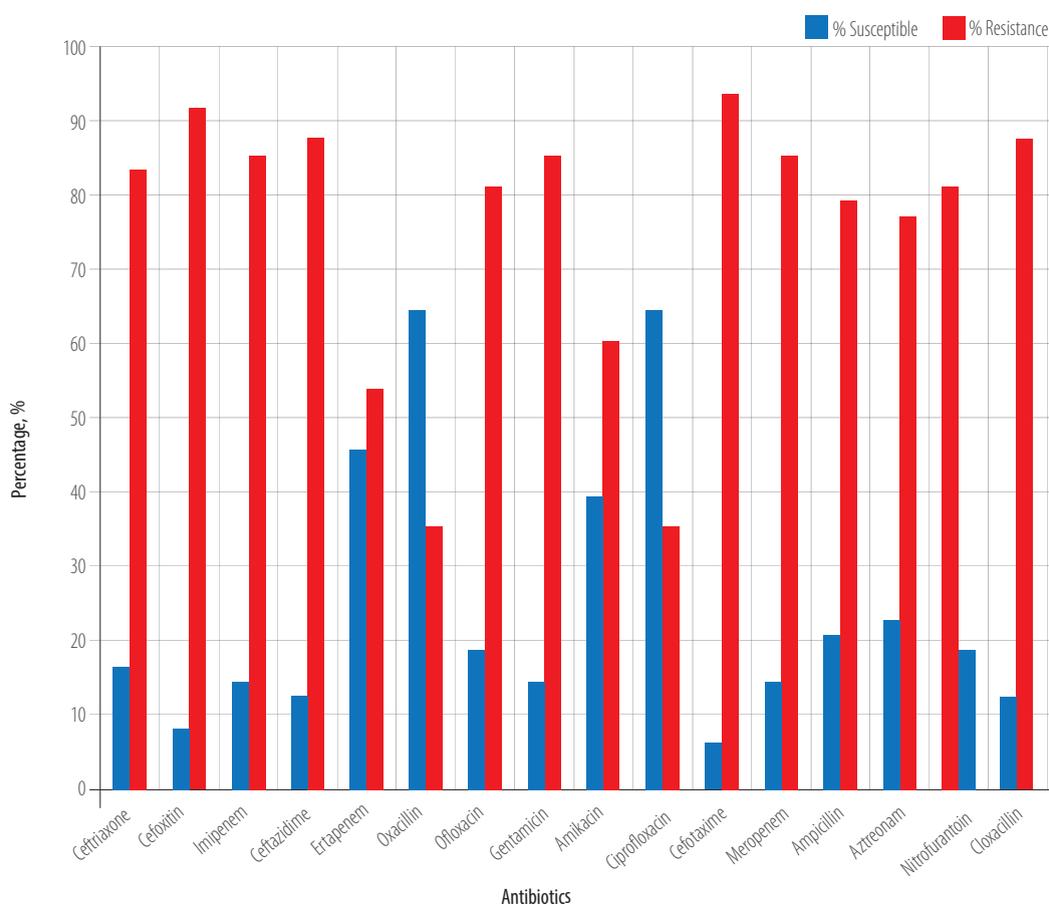


Figure 1: Susceptibility profile of the isolated *Escherichia coli*.

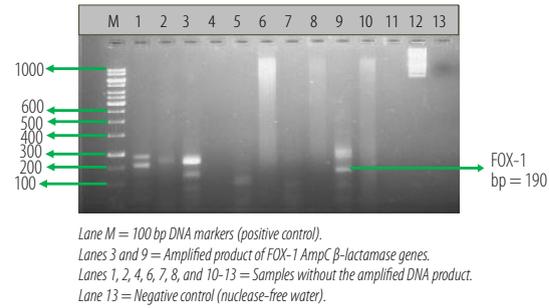
Table 2: Frequency of AmpC-producing *Escherichia coli* isolates.

Organism	Total no.	CC-DDST*	FOX-1 positive	AmpC genes negative
<i>E. coli</i> , n (%)	48	6 (12.5)	3 (6.3)	45 (93.8)

*Cefoxitin-cloxacillin double-disk synergy test (CC-DDST).

cefoxitin disk used alone was indicative of AmpC enzyme production phenotypically.

The presence of AmpC β -lactamase genes in the *E. coli* isolates was investigated using a multiplex PCR assay targeting the FOX-1 AmpC β -lactamase genes as described by Perez-Perez and Hanson.¹⁵ Each of the *E. coli* isolates was inoculated from nutrient agar plates (Oxoid, UK) into 5 mL of peptone water (Oxoid, UK). The inoculated broths were incubated at 30 °C for 18–24 hours with mild shaking. Bacterial cells from the overnight broth culture were aseptically harvested by centrifugation at 5000 rpm for 5 minutes, and the supernatant was decanted. The pellets collected at the bottom of the tube were resuspended in 100 μ L of Tris-EDTA (TE) buffer. The cells in the TE buffer tubes were lysed by heating at 100 °C for 10 minutes in a water bath. Finally, the solution was centrifuged at 5000 rpm for 5 minutes (to remove cellular debris). The supernatant was used as the DNA template source for gene amplification. Multiplex PCR amplification of AmpC β -lactamase genes from the test isolates was carried out using primers synthesized and supplied by Inqaba Biotechnical Industries Ltd (Inqaba Biotechnical Industries Ltd, South Africa) in a thermal cycler (Lumex Instruments, Canada) and with a final PCR mixture of 26.5 μ L containing 0.2 μ L of Taq polymerase enzyme U/ μ L, 2.5 μ L of 10X PCR buffer along with 2.5 μ L MgCl₂, 1 μ L of 10 pM from each of the forward and reverse primers, 2.5 μ L of dNTPs MIX (2 Mm), 3 μ L of DNA template (from the test isolates), and 14.8 μ L of nuclease-free water. To determine the *E. coli* isolates possessing the FOX-1 AmpC β -lactamase genes, primers for FOX-1 genes were used: FOXM-Forward (AAC ATG GGG TAT CAG GGA GAT G) and FOXM-Reverse (CAA AGC GCG TAA CCG GAT TGG).¹⁵ Amplification included initial denaturation at 94 °C for 3 minutes, followed by 25 cycles of DNA denaturation at 94 °C for 30 seconds, annealing at 64 °C for 30 seconds, and extension at 72 °C for

**Figure 3:** Electrophoretogram showing the FOX-1 AmpC β -lactamase genes amplified from two different *Escherichia coli* isolates.

1 minute. Gel electrophoresis of the multiplex PCR products was carried out using 1.5% agarose gel (Inqaba Biotechnical Industries Ltd, South Africa) for 2 hours at 80 V.

RESULTS

A total of 120 swab samples were aseptically collected from the anal region of cows in a slaughterhouse in Abakaliki, Nigeria. *E. coli* was isolated from 48 swab samples giving a rate of 40.0% [Table 1].

β -lactamase enzyme production was detected in 35 (72.9%) isolates of *E. coli* [Table 1]. We observed a high rate of resistance among the test *E. coli* isolates. The *E. coli* isolates were highly resistant to the carbapenems, penicillins, aminoglycosides, fluoroquinolones, and cephalosporins [Figure 1]. Overall, the percentage resistance of the *E. coli* isolates to the tested antibiotics shows that the organism was resistant to cefoxitin (91.7%), ceftriaxone (83.3%), imipenem (85.4%), ceftazidime (87.5%), ofloxacin (81.3%), and gentamicin (85.4%). However, cefotaxime, meropenem, ampicillin, aztreonam, and cloxacillin also showed the least susceptibility to the tested isolates at the rate of 93.7%, 85.4%, 79.2%, 77.1%, and 87.5%, respectively [Figure 2].

AmpC β -lactamase production was also detected phenotypically in six *E. coli* (12.5%) isolates using the CC-DDST. Table 2 also shows the result of the molecular detection of plasmid-mediated FOX-1 AmpC β -lactamase genes in the *E. coli* isolates by multiplex PCR. Among the 48 *E. coli* isolates used for this study, only three harbored the FOX-1 plasmid-mediated AmpC gene, which provokes the production of AmpC β -lactamase in the test organism. A total of 45 *E. coli* isolates

showed no AmpC gene [Table 2]. Electrophoretic analysis of the PCR product of the test isolates after gene amplification revealed bands in the electrophoretogram, and this was positive for FOX-1 gene with a base pair size of 190 bp [Figure 3].

DISCUSSION

The use of antibiotics in animal husbandry and other agricultural practices has contributed significantly to the burden and threat of antimicrobial resistance in the community. The improper handling and usage of animal wastes and effluents from abattoirs (inundated with antibiotic-resistant bacteria) in this part of the world necessitated our study, which investigated phenotypically and genotypically the prevalence, antibiogram, and genetic determinants of AmpC-producing *E. coli* from a local abattoir in Abakaliki metropolis, Ebonyi State, Nigeria. The occurrence of gram-negative bacteria including *E. coli* in environmental samples has been previously reported within Nigeria and in other parts of the world.^{11,12,16,17} Another study from Nigeria also reported that *Enterobacteriaceae* were the most prevalent organisms isolated from environmental samples.¹¹ Several studies have also shown *E. coli* was the most prevalent gram-negative bacteria isolated from environmental samples including samples from poultry birds.^{16–20} The presence of β -lactamase enzyme in bacteria provides opportunity for the horizontal transmission of these enzymes from one organism to another.^{1,16,17,21–23} The high levels of resistance of *E. coli* isolates from environmental samples to some conventional antibiotics (as obtainable in this study) have been reported in the Netherlands, Nigeria, and Uganda.^{24–27} Bergenholtz et al,¹⁷ also reported high resistance of *E. coli* isolates from environmental samples to some conventional antibiotics as reported in this study. In Uganda, it was reported that 168 *E. coli* isolates out of 182 were resistant to several antibiotic classes including penicillins, fluoroquinolones, and aminoglycosides.²⁵

Our study revealed that AmpC enzyme production was high in anal swab samples. In India, the prevalence of AmpC-producing *E. coli* isolates was 12.5%, a result similar to ours.²⁸ However, the prevalence of AmpC enzyme-producing *E. coli* isolates in Iran was reported as 37.2%,²⁹ and this result is higher than the occurrence rate of AmpC

enzyme-producing *E. coli* isolates in this study. In the Czech Republic, two *E. coli* isolates from cloacal swabs of poultry birds were reported as AmpC-positive.³⁰ The FOX gene homologs are usually the most prevalent AmpC genes aside from the CMY gene homologs found in gram-negative bacteria, which mediate their resistance to the cephamycins.^{3,31} The prevalence of *E. coli* phenotypes harboring the FOX-1 gene in this study is not in agreement with a study from Egypt, which detected the FOX-1 gene in 33.3% *E. coli* isolates.³² We detected the FOX-1 genes in only 6.3% of *E. coli* isolates. Another study from Egypt reported AmpC-producing strains in 88.46% isolates among cefoxitin resistant *E. coli* and *Klebsiella* species.³³ The low percentage of FOX family-specific AmpC genes detected by the authors is in accordance with our study, which we detected by multiplex PCR.³³ In Iran, the prevalence of AmpC-producing strains of *E. coli* was 37.2%, a result higher than ours.²⁹ In another related study from Pakistan, it was also reported that the FOX genes were the most prevalent AmpC genes detected by PCR in *E. coli* isolates.³⁴

The prevalence, emergence and spread of antibiotic-resistant bacteria in both the community and hospital environment as previously reported is notable and can impact negatively on antimicrobial therapy.^{33–36} In a related and recent study, it was reported that the antimicrobial armamentarium might soon be depleted due to the nefarious activities of antibiotic-resistant bacteria which have put the therapeutic effect of some drugs at risk.³⁵

CONCLUSION

We report the first multiplex detection of FOX-1 AmpC genes in gram-negative bacteria in Abakaliki, Nigeria. Our results show that *E. coli* isolates from abattoir origin harbor antibiotic resistance genes (specifically AmpC genes) that mediate gram-negative resistance to the cephamycins. Minimizing the use of broad-spectrum antibiotics in both clinical and non-clinical settings will help to contain the emergence and spread of these organisms. The use of antibiotics to treat animals and as growth promoting agents should be curtailed and possibly replaced by proper hygienic practices and the use of vaccination to limit the emergence and spread of antibiotic-resistant bacteria in the community.

Disclosure

The authors declared no conflicts of interest. No funding was received for this study.

Acknowledgements

We appreciate the members and staff of the Biotechnology Research and Development Center of Ebonyi State University, Abakaliki for their technical guidance in carrying out the multiplex PCR technique.

REFERENCES

- Bush K, Jacoby GA. Updated functional classification of β -lactamases. *Antimicrob Agents Chemother* 2010 Mar;54(3):969-976.
- Bush K, Jacoby GA, Medeiros AA. A functional classification scheme for beta-lactamases and its correlation with molecular structure. *Antimicrob Agents Chemother* 1995 Jun;39(6):1211-1233.
- Walsh TR, Toleman MA, Poirel L, Nordmann P. Metallo- β -lactamases: the quiet before the storm? *Clin Microbiol Rev* 2005 Apr;18(2):306-325.
- Shevade SU, Agrawal GN. A study on the AmpC production amongst the urinary Enterobacteriaceae isolates. *J Clin Diagn Res* 2013 Aug;7(8):1831-1832.
- Vanwynsberghe T, Verhamme K, Raymaekers M, Cartuyvels R, Vaerenbergh KV, Boel A, et al. A Large hospital outbreak of *Klebsiella pneumoniae* (DHA-1 and SHV-11 Positive): Importance of detection and treatment of AmpC β -Lactamases. *The Open Infectious Diseases Journal* 2009;3:55-60.
- Jacoby GA, Munoz-Price LS. The new β -lactamases. *N Engl J Med* 2005 Jan;352(4):380-391.
- Fernández M, Conde S, de la Torre J, Molina-Santiago C, Ramos JL, Duque E. Mechanisms of resistance to chloramphenicol in *Pseudomonas putida* KT2440. *Antimicrob Agents Chemother* 2012 Feb;56(2):1001-1009.
- Cheesbrough M. *District Laboratory Practice in Tropical Countries*. 2nd ed. Cambridge University Press, UK; 2006. p. 178-187.
- Javeed I, Hafeez R, Anwar MS. Antibiotic susceptibility pattern of bacterial isolates from patients admitted to a tertiary care hospital in Lahore. *Biomedica* 2011;27:19-23.
- Clinical and Laboratory Standard Institute. Performance standards for antimicrobial disk susceptibility testing. Fifteenth informational supplement, CLSI document M100-S15. USA: Wayne, PA; 2011.
- Akinduti PA, Ejilude O, Motayo BO, Adeyokinu AF. Emerging multidrug resistant AmpC beta-lactamase and carbapenemase enteric isolates in Abeokuta, Nigeria. *Nat Sci* 2012;10(7):70-74.
- Ejikeugwu C, Esimone C, Iroha I, Ugwu C, Ezeador C, Duru C, et al. Phenotypic detection of AmpC beta-lactamase among anal *Pseudomonas aeruginosa* isolates in a Nigerian abattoir. *Archives of Clinical Microbiology* 2016;7(2):1-5.
- Manchanda V, Singh NP, Shamweel A, Eideh HK, Thukral SS. Molecular epidemiology of clinical isolates of ampc producing *Klebsiella pneumoniae*. *Indian J Med Microbiol* 2006 Jul;24(3):177-181.
- Polsfuss S, Bloemberg GV, Giger J, Meyer V, Böttger EC, Hombach M. Practical approach for reliable detection of AmpC beta-lactamase-producing Enterobacteriaceae. *J Clin Microbiol* 2011 Aug;49(8):2798-2803.
- Pérez-Pérez FJ, Hanson ND. Detection of plasmid-mediated AmpC β -lactamase genes in clinical isolates by using multiplex PCR. *J Clin Microbiol* 2002 Jun;40(6):2153-2162.
- Ben Slama K, Jouini A, Ben Sallem R, Somalo S, Sáenz Y, Estepa V, et al. Prevalence of broad-spectrum cephalosporin-resistant *Escherichia coli* isolates in food samples in Tunisia, and characterization of integrons and antimicrobial resistance mechanisms implicated. *Int J Food Microbiol* 2010 Feb;137(2-3):281-286.
- Bergenholtz RD, Jørgensen MS, Hansen LH, Jensen LB, Hasman H. Characterization of genetic determinants of extended-spectrum cephalosporinases (ESCs) in *Escherichia coli* isolates from Danish and imported poultry meat. *J Antimicrob Chemother* 2009 Jul;64(1):207-209.
- Bashir D, Thokar MA, Fomda BA, Bashir G, Zahoor D, Ahmad S, et al. Detection of metallo- β -lactamase (MBL) producing *Pseudomonas aeruginosa* at a tertiary care hospital in Kashmir. *Afr J Microbiol Res* 2011;5(2):164-172.
- Chakraborty D, Basu S, Das S. A study on infections caused by Metallo-Beta-Lactamase Producing Gram-negative Bacteria in Intensive Care Unit Patients. *Am J Infect Dis* 2010;6(2):34-39.
- Leung GH, Gray TJ, Cheong EY, Haertsch P, Gottlieb T. Persistence of related bla-IMP-4 metallo-beta-lactamase producing Enterobacteriaceae from clinical and environmental specimens within a burns unit in Australia - a six-year retrospective study. *Antimicrob Resist Infect Control* 2013 Dec;2(1):35.
- Aibinu I, Nwanneka T, Odugbemi T. Occurrence of ESBLs and MBL in Clinical Isolates of *Pseudomonas aeruginosa* from Lagos, Nigeria. *J Am Sci* 2007;3(4):81-85.
- Akujobi CO, Odu NN, Okorundu SI. Detection of AmpC beta lactamases in clinical isolates of *Escherichia coli* and *Klebsiella*. *Afr J Clin Exp Microbiol* 2012;13(1):51-55.
- Usha PT, Sabitha J, Nisha AR. Antimicrobial drug resistance: A global concern. *Vet World* 2010;3(3):138-139.
- van den Bogaard AE, London N, Driessen C, Stobberingh EE. Antibiotic resistance of faecal *Escherichia coli* in poultry, poultry farmers and poultry slaughterers. *J Antimicrob Chemother* 2001 Jun;47(6):763-771.
- Majalija S, Francis O, Sarah WG, Lubowa M. Antibiotic susceptibility profiles of fecal *Escherichia coli* isolates from dip-litter broiler chickens in Northern and Central Uganda. *Vet Res* 2010;3(4):75-80.
- Ogunleye AO, Oyekunle MA, Sonibare AO. Multidrug resistant *Escherichia coli* isolates of poultry origin in Abeokuta, South Western Nigeria. *Vet Arh* 2008;78(6):501-509.
- European Food Safety Authority (EFSA). EFSA panel on biological hazards; scientific opinion on the public health risks of bacterial strains producing extended-spectrum beta-lactamases and/or AmpC beta-lactamase in food and food-producing animals. *EFSA J* 2011;9(8):2322.
- Manoharan A, Sugumar M, Kumar A, Jose H, Mathai D, Khilnani GC, et al. ICMR-ESBL group. Phenotypic & molecular characterization of AmpC β -lactamases among *Escherichia coli*, *Klebsiella* spp. & *Enterobacter* spp. from five Indian Medical Centers. *Indian J Med Res* 2012 Mar;135:359-364.
- Mansouri S, Kalantar Neyestanaki D, Shokoohi M, Halimi S, Beigverdi R, Rezagholezadeh F, et al. Characterization of AmpC, CTX-M and MBLs types of β -lactamases in clinical isolates of *Klebsiella pneumoniae* and *Escherichia coli* producing Extended Spectrum β -lactamases in Kerman, Iran. *Jundishapur J Microbiol* 2014 Feb;7(2):e8756.
- Kolar M, Bardon J, Chroma M, Hricova K, Stosova T, Sauer P, et al. ESBL and AmpC beta-lactamase-producing Enterobacteriaceae in poultry in the Czech Republic. *Vet Med (Praha)* 2010;55(3):119-124.
- Thomson KS. Extended-spectrum- β -lactamase, AmpC, and Carbapenemase issues. *J Clin Microbiol* 2010 Apr;48(4):1019-1025.
- El-Hady SA, Adel LA. Occurrence and detection of AmpC β -lactamases among Enterobacteriaceae isolates from patients at Ain Shams University Hospital. *Egypt J Med Hum Genet* 2015;16:239-244.
- Helmy MM, Wasfi R. Phenotypic and molecular

- characterization of plasmid mediated AmpC β -lactamases among *Escherichia coli*, *Klebsiella* spp., and *Proteus mirabilis* isolated from urinary tract infections in Egyptian hospitals. Biomed Research International, Hindawi Publishing Corporation 2014. p. 1-9.
34. Hussain M, Hasan F, Shah AA, Hameed A, Jung M, Rayamajhi N, et al. Prevalence of class A and AmpC β -lactamases in clinical *Escherichia coli* isolates from Pakistan Institute of Medical Science, Islamabad, Pakistan. Jpn J Infect Dis 2011;64(3):249-252.
 35. Balkhair A. Antibiotics Resistance: When the Armamentarium Gets to the Verge of Being Empty. Oman Med J 2017 Jul;32(4):267-268.
 36. Zorgani A, Almagatef A, Sufya N, Bashein A, Tubbal A. Detection of CTX-M-15 Among Uropathogenic *Escherichia coli* Isolated from Five Major Hospitals in Tripoli, Libya. Oman Med J 2017 Jul;32(4):322-327.